PASC in Pregnancy and Postpartum Registry | Biospecimens Recommended Measures

Tier 1 Biospecimens (Neonate)					
Specimen	Collection	Timing	Storage	Sample Analyses	
Neonate/Cord Blood	Plasma / Buffy Coat: EDTA tubes (at least 10mL total) Serum: Serum separator (5mL)	Date of Draw	 Plasma: Freeze in 200 μL (max 1 mL) aliquots, store at -80°C Serum: Freeze in200 μL (max 1 mL) aliquots, store at -80°C Buffy coat: Freeze, store at -80°C 	Plasma / Serum: SARS-CoV-2 RNA viral load SARS-CoV-2 antibodies (IgG, IgA, neutralizing antibody evaluation) Cytokine analysis Buffy coat: Analysis of cellular fraction (Evaluation of properties of specific cellular fractions such as T-cells, monocytes)	
Neonatal Respiratory Specimens	 Nasopharyngeal swab (preferred) Nasal swab Oropharyngeal swab (RTq-PCR) Saliva vial / cup 	 Delivery At time of acute illness (if applicable)¹⁸ 	 Swab: Swirl in PBS, aliquot PBS and freeze at -80°C If diagnostic swab can be retrieved from clinical lab, it can be used for quantitative (viral load) and other assays Saliva: Process with DTT, store at -80C 	Use to confirm SARS-CoV-2 negative status in control group SARS-CoV-2 RNA viral load	

	Tier 1 Biospecimens (Maternal)					
Specimen	Collection	Timing	Storage	Sample Analyses		
Maternal Blood ¹⁹	Plasma / Buffy Coat: EDTA tubes (at least 10mL total) Serum: Serum separator (5mL)	 At time of acute illness and / or At delivery 	 Plasma: Freeze in 200 μL (max 1 mL) aliquots, store at -80°C Serum: Freeze in200 μL (max 1 mL) aliquots, store at -80°C Buffy coat: Freeze, store at -80°C 	Plasma / Serum: SARS-CoV-2 RNA viral load SARS-CoV-2 antibodies (IgG, IgA, neutralizing antibody evaluation) Cytokine analysis Buffy coat: Analysis of cellular fraction (Evaluation of properties of specific cellular fractions such as T-cells, monocytes)		
Cord Blood	 Plasma / Buffy Coat: EDTA tubes (at least 10-15 mL total, may be less for pre-term deliveries) Serum: Serum separator (7.5mL) 	At delivery	 Plasma: Freeze in 200 μL (max 1 mL) aliquots, store at -80°C Serum: Freeze in 200 μL (max 1 mL) aliquots, store at -80°C Buffy coat: Freeze, store at -80°C 	 Plasma / Serum: SARS-CoV-2 RNA viral load SARS-CoV-2 antibodies (IgG, IgA, neutralizing antibody evaluation) Cytokine analysis Buffy coat: Analysis of cellular fraction (Evaluation of properties of specific cellular fractions such as T-cells, monocytes) 		
Placenta	Fixed tissue	At delivery	 Store at 4°C until fixation Once fixed, can be stored as formalin-fixed paraffin embedded blocks 	• RNA <i>in situ</i> hybridization (RNA-ISH)		
	Maternal side biopsy andFetal side biopsy	 As soon as possible; within 1-2 hours of delivery maximum (RNA will degrade) 	 Process in RNA later, following manufacturer instructions Store preserved tissue at -80°C (or -20°C if 80°C not available) 	• SARS-CoV-2 RNA analyses ²⁰		
Colostrum and/or mature milk	 Pump into colostrum cup or Hand expression into colostrum cup For larger volume: Pump into pumping containers or storage containers 	 During delivery admission or At postpartum visit 	Small volume: Aliquot (e.g. 1 mL aliquots) and store at -80°C Large volume (e.g. 10 mL or above): Spin and separate cellular fraction and supernatant. Aliquot (e.g. 1 mL aliquots) and store at -80°C	 SARS-CoV-2 RNA viral load²¹ SARS-CoV-2 antibodies (IgG, IgA, neutralizing antibody evaluation) 		

Tier 2 Expanded Specimen Collection (Maternal)				
Specimen	Collection	Timing	Storage	Sample Analyses
Maternal	 Nasopharyngeal 	• 11-14 weeks ²³	• Swab: Swirl in PBS, aliquot PBS	• Use to confirm SARS-CoV-2
Respiratory	swab (preferred)	• 18-22 weeks ⁵	and freeze at -80°C	negative status in control
Specimens ²²	 Nasal swab 	• 28-32 weeks ⁵		group

	 Oropharyngeal swab (RTq-PCR) Saliva vial / cup 	 Delivery At time of acute illness (if applicable)²⁴ 	 If diagnostic swab can be retrieved from clinical lab, it can be used for quantitative (viral load) and other assays Saliva: Process with DTT, store at -80C 	SARS-CoV-2 RNA viral load
Maternal Blood	Plasma, Buffy Coat, and/or PBMC: EDTA tubes (at least 10mL total) Serum: Serum separator (5mL)	 11-14 weeks⁵ 18-22 weeks⁵ 28-32 weeks⁵ Delivery At time of acute illness 	 Plasma: Freeze in 200 μL (max 1 mL) aliquots, store at -80°C Serum: Freeze in 200 μL (max 1 mL) aliquots, store at -80°C Buffy coat: Freeze, store at -80°C PBMC: Store in freezing media in liquid nitrogen (LN₂) 	 Inflammatory markers (e.g. IL-6, TNF-a, IL-1B, IFN-g, IL-10, CRP/ESR) T-cell, monocyte, other specific cell fraction experiments (EDTA only) AM cortisol or CRH
	PaxGene tube (2.5 or 5 mL)	 11-14 weeks⁵ 18-22 weeks⁵ 28-32 weeks⁵ Delivery At time of acute illness 	 Shake vigorously May keep at room temp for 2- 24 hrs Freeze whole tube, store at - 80°C 	Transcriptomic/global gene expression analyses
Cord Blood	• For PBMC: EDTA tubes (at least 10-15 mL total)	At delivery	• Isolated PBMC: Store in liquid nitrogen (LN ₂)	Analysis of cellular fraction (scRNA-Seq, evaluation of properties of specific cellular fractions such as T-cells, monocytes) Genotyping (array-based)
	PaxGene tube (2.5 or 5 mL)	At delivery	 Shake vigorously Keep at room temp for 2-24 hrs Freeze whole, store at -80°C 	Transcriptomic/global gene expression analyses
Placenta	 Maternal side placental biopsy and Fetal side placental biopsy 	As soon as possible; within 1-2 hours of delivery maximum (RNA will degrade)	 Process in RNA later, following manufacturer instructions Store preserved tissue at -80°C 	 SARS-CoV-2 RNA viral load RNA RTq-PCR for specific genes of interest DNA methylation analyses Genotyping (array-based)
	 Full thickness biopsies or Remaining whole placenta 	Take biopsies in pathology lab	Formalin-fixed, paraffin- embedded blocks	RNA in situ hybridization (RNA-ISH) to define placental infection

¹ If a participant that was previously designated as a control becomes ill with COVID-19, specimens collected after that point can no longer be used as a control

¹⁹ Blood draws for research in pregnancy should not exceed 50 mL in 8 weeks. Those with Hct < 24 should not provide blood for research

²⁰ Preservation in RNA later permits both RNA and DNA analyses

²¹ Specific guidance to participants and adherence to breast cleaning protocols is critically important if breastmilk viral load quantification is planned

²² Respiratory specimens only necessary to be collected to document COVID-19 negative status at the time of collection if the participant provides other samples for study

²³ Can alternatively collect at 1st, 2nd, 3rd trimester appointments. Ideally, align maternal respiratory specimens collection with maternal blood draw ²⁴ If a participant that was previously designated as a control becomes ill with COVID-19, specimens collected after that point can no longer be used as a control

			Fixed tissue can be sectioned and stored on slides and in in paraffin-embedded blocks	 ACE2/TMPRSS2 protein expression patterns Cd68/CD163 for Hofbauer cell hyperplasia and chronic histiocytic intervillositis
Breastmilk	Large volume:Pump into pumping containers or storage containers	At delivery and/orPost-partum	Large volume (e.g. 10 mL or above): Spin and separate cellular fraction and supernatant	Cellular fraction of breastmilk: Sequencing Isolation of T-cells, NK cells, and antibody-producing B cells

Tier 3 (Maternal)				
Specimen	Collection Method	Timing	Storage	Sample Analyses
Saliva (Tier 3)	Oragene	• Follow time restrictions included in instructions	 Process with DTT Store at -80°C 	Genotyping
Placenta	Maternal biopsy andFetal side biopsy	 As soon as possible; within 1-2 hours of delivery maximum (RNA will degrade) 	 Snap freeze tissue in liquid nitrogen (preferred) or on dry ice Store at -80°C 	Protein isolationSingle-cell RNA-seqDNA/RNA extraction
	Full thickness biopsies orRemaining whole placenta	Take biopsies in pathology lab	 Formalin-fixed, paraffinembedded blocks Fixed tissue can be sectioned and stored on slides and in in paraffin-embedded blocks 	CD147 & CD26 protein expression patterns
	Membrane or decidua basalis	 As soon as possible Within 1-2 hours of delivery maximum (RNA will degrade) 	Process fresh for cell isolation per protocol	Inflammatory/immune analyses (e.g., FACS, flow cytometry, transcriptomics, pro-inflammatory cytokine quantification)